

Central European Institute of Technology BRNO | CZECH REPUBLIC

Jan Přibyl

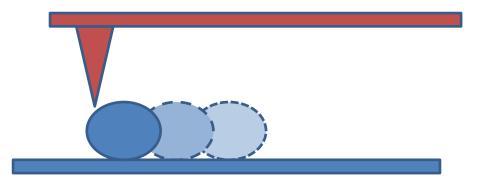
Core Facility NanoBiotechnology, CEITEC MU Masaryk university, Brno, Czech Republic *E-mail: jan.pribyl@ceitec.muni.cz* 

### Specific approaches in microscopy samples preparation

Multimodal Microscopy Workshop: Probing the Triad of Structure, Mechanics, and Chemistry in Biological Systems, 2024, Brno 

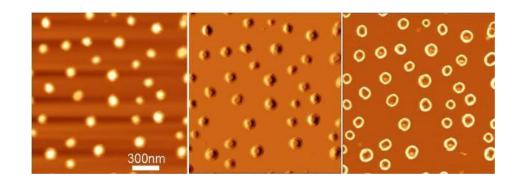
# Sample preparation for AFM

# **AFM sample preparation**

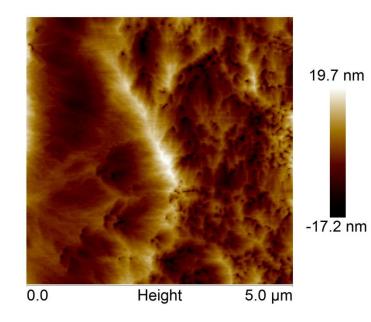




#### **Concentration – surface density**



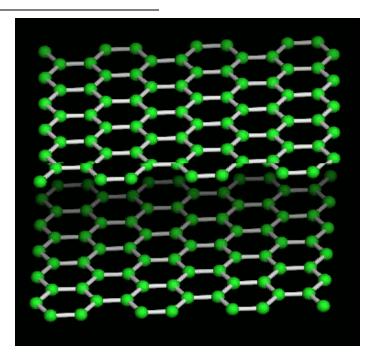




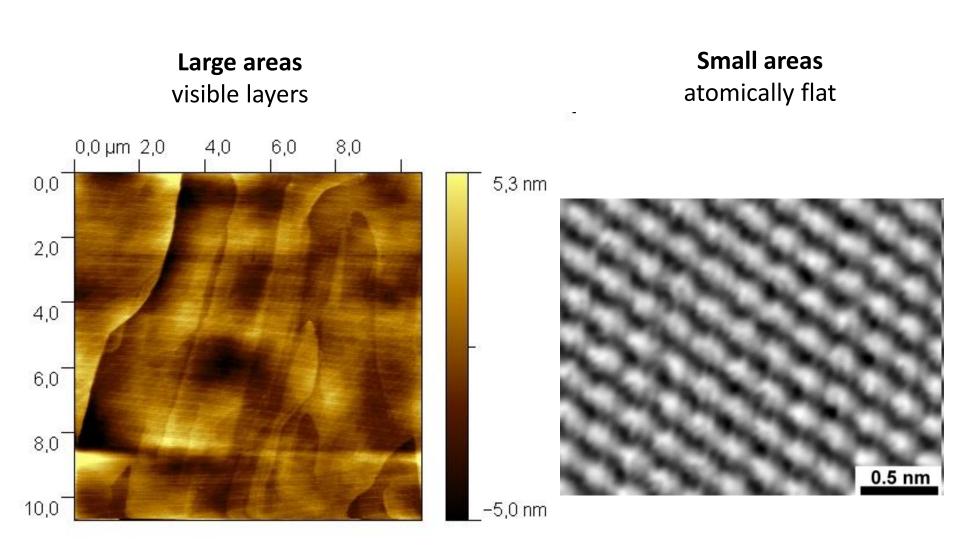
**Substrates** for preparation of AFM samples

#### 1. HOPG Highly Ordered Pyrolytic Graphite

- •Kish's graphite, waste in steel production
- •Hexagonal planar structure
- •C-C bond142 pm, layer-layer distance335 pm
- Conductive, highly hydrophobic
- Planar structure
- •Synthetic form of graphite, high chemical purity
- Traditionally substrate for SEM, STM i AFM (→ conductivity)
- Immobilization spontaneous adsorption (→ hydrophobicity)

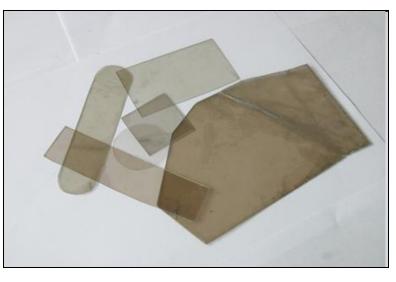


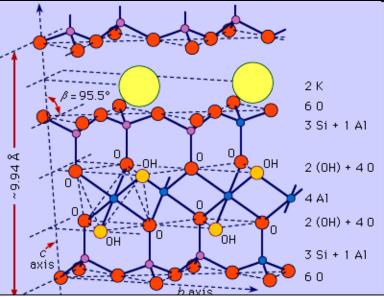
1. HOPG Highly Ordered Pyrolytic Graphite



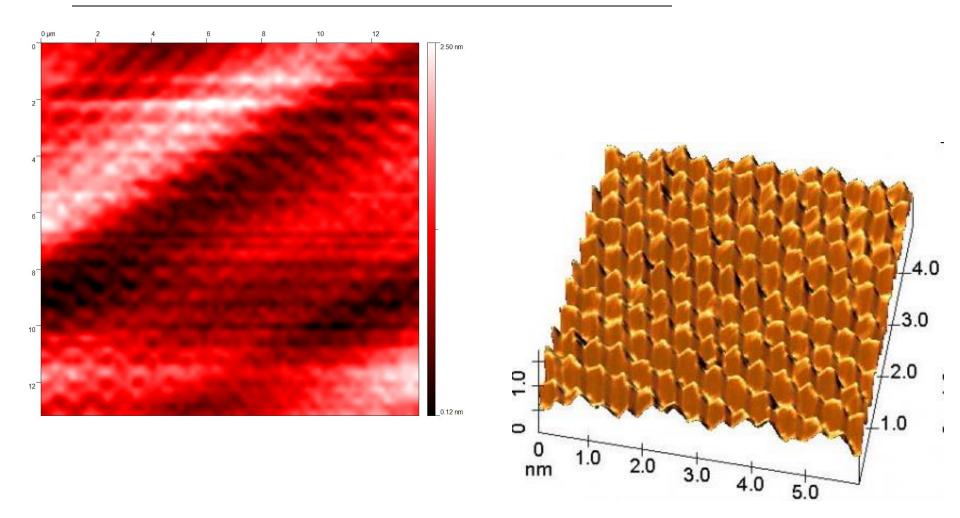
2. Mica (muscovite)

- "Cat's silver", muscovite acc. to city of Moscow
- •Chem. structure:  $K_2O \cdot Al_2O_3 \cdot SiO_2$
- •Hydrophilic surface
- Easy to be modified by chemical synthesis
- Immobilization by chemical bonding as well as ionic interaction
- •pKa ~ 3, physiological pH → negative surface charge
- Mica = silicate, hydrated SiO<sub>2</sub> (~ Si-OH) from the chemical point of view





#### 2. Mica (muscovite)

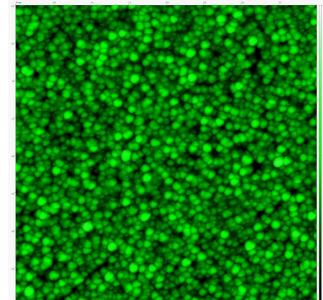


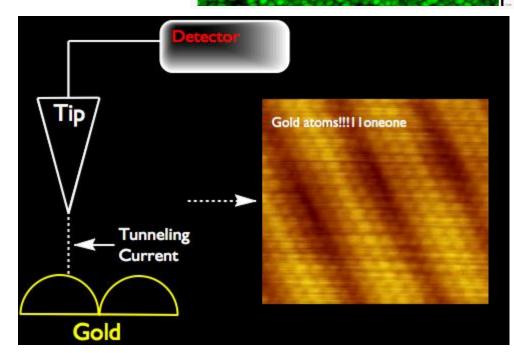
**Extremely flat on small and larger areas** 

### Other surfaces 3. Gold

- Inert metal
- •Traditionally in (bio)electrochemistry (i.e. biosensors) electrodes
- •Conductive STM + AFM
- Hydrophobic: spontaneous non-selective adsorption of molecules (proteins, DNA, ...)
- •Specific chemical binding of thiols (-SH) – organic molecules + cysteine
- Prepared usually by evaporation
- •Adhesion layer for operation in liquids (Al/Cr/Ti)

**Sputtered gold layer** image by tapping mode AFM





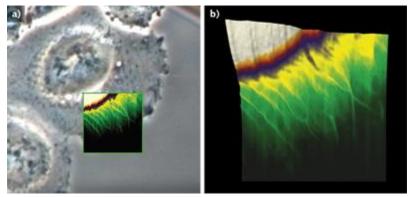
# **Other surfaces**

#### 4. Glass

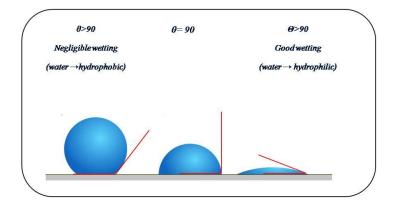
Amorphous noncrystalline structure



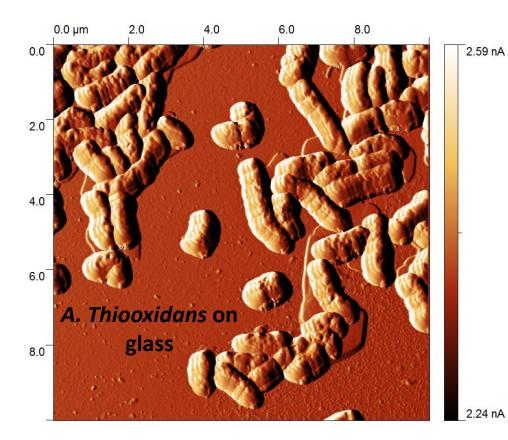
- •Lab glass composition: 75% SiO<sub>2</sub> plus Na<sub>2</sub>O, CaO, borate and minor additives
- •Si-OH  $\rightarrow$  from chemical point of view
- Less hydrophilic comparing to mica
- •Roughness much higher comparing to mica (production by pressing)
- •Not suitable for individual molecules imaging with AFM
- •Typically used together with optical microscopy cell compartments, whole cells

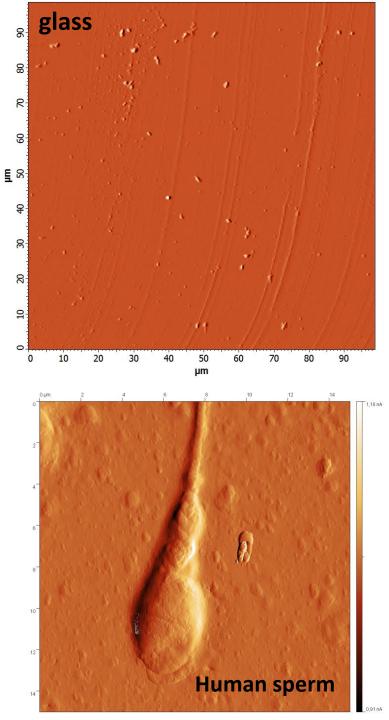


AFM – optical image overlap



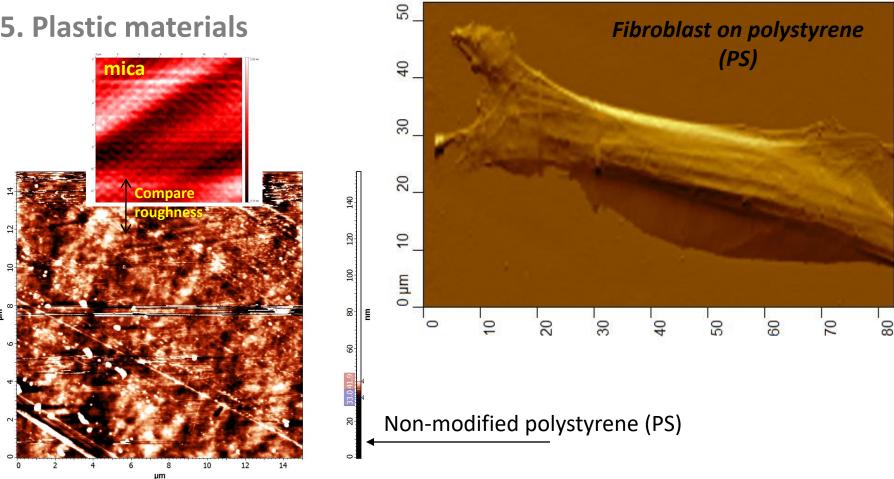
#### Whole cells on glass under AFM





# **Other surfaces**





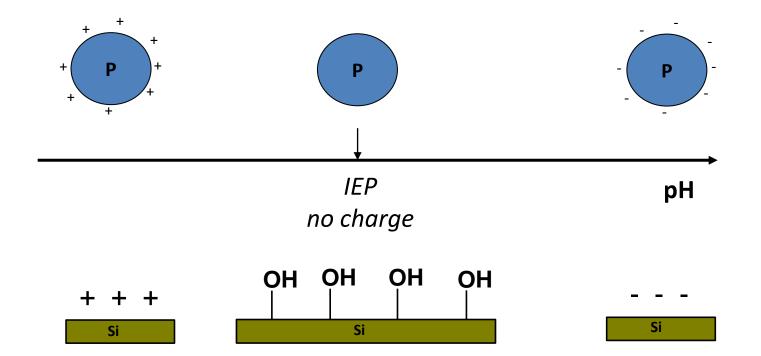
- Most of lab supplies made of plastic (PP, PE, PS)
- No functional groups to be used in covalent binding
- PS hydrophobic  $\rightarrow$  spontaneous non-specific adsorption of proteins  $\rightarrow$  usually as underlying support (i.e. for cell attachment)

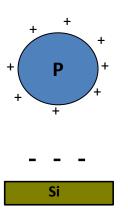
# **Immobilization procedures**

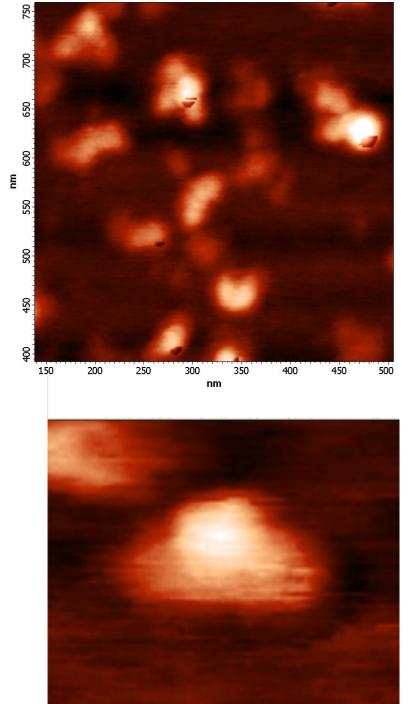
#### **1. Proteins**

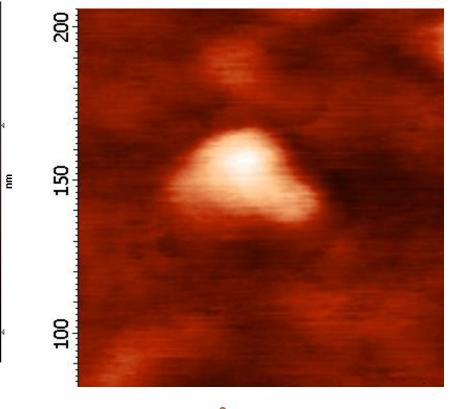
Surface: mica or HOPG (extremely flat)

Protein: charge is given by IEP + pH
Immobilization on mica: pKa (mica) < pH < IEP</pre>









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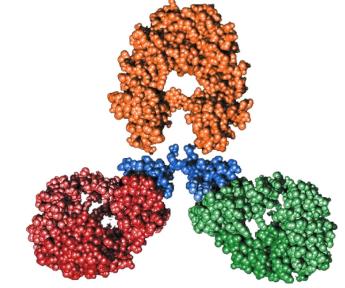
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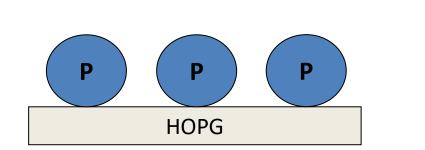
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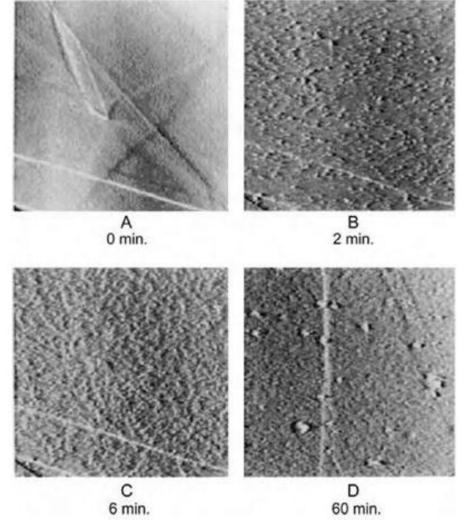


#### **Protein immobilization on HOPG**

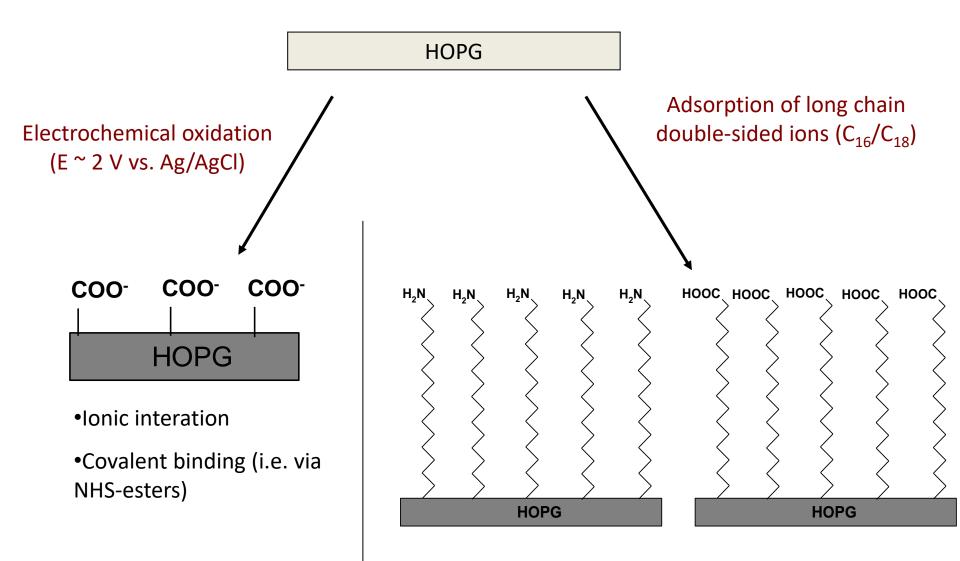
A. **Spontaneous** (non-specific) **adsorption** of protein  $\rightarrow$  hydrophobic surface (best results at zero charge *pH* = *IEP*)



Lysozyme molecules on HOPG

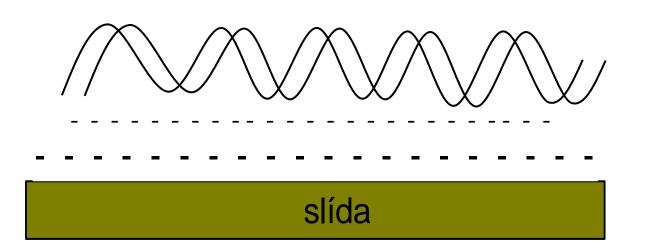


B. **Ionic** (specific) **binding** of molecules  $\rightarrow$  creation of charge/chem. groups on HOPG surface



#### **2. DNA**

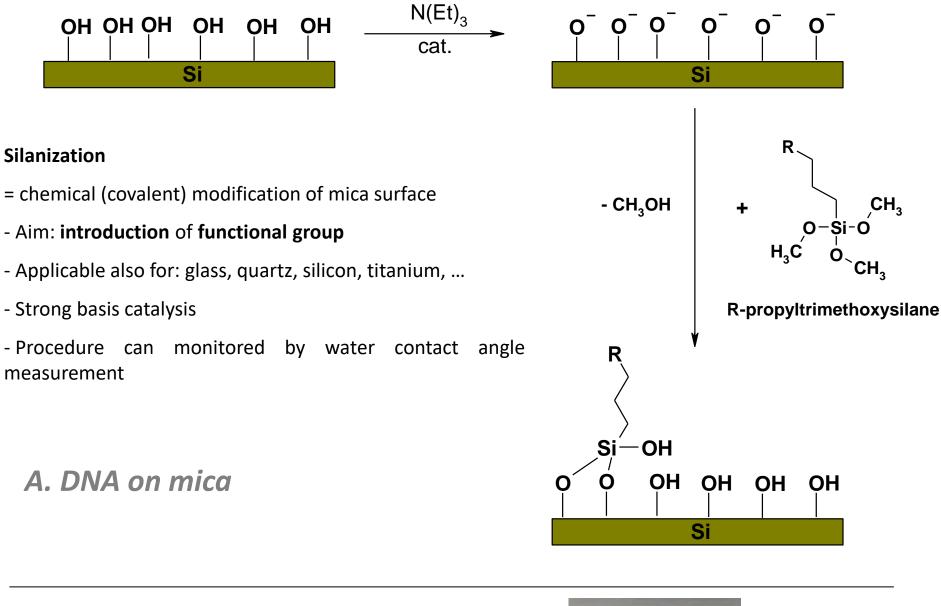
Surface: mica or HOPG (extremely flat)



#### Immobilization problem:

**DNA** (sugar-phosphate bone) as well as **mica – negative charge** under physiological pH

 $\rightarrow~$  surface introduction of **positive charge** 



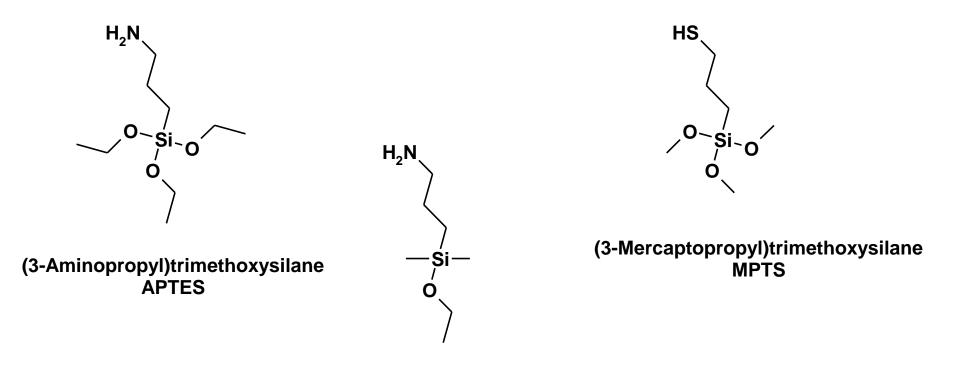


silanization

hydrophobization



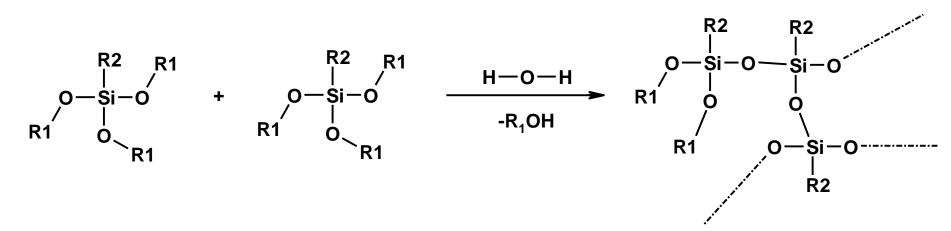
#### Examples of alkoxysiloxanes



3-(Ethoxydimethylsilyl)propylamine APDMES

### **Self-polymerization**

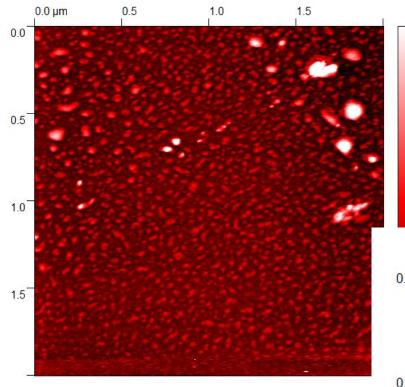
practical complication



- Especially with **APTES** during liquid silanization
- Even vapors of water can cause this effect
- Fixation for **optical** microscopy **expected** factor
- In contrary in fixation for **AFM** very **disturbing**
- Solution:
  - silanization in **vapours** under **vacuum** (i.e. in desiccator<mark>s)</mark>
  - monoalkoxysilanes can not polymerize



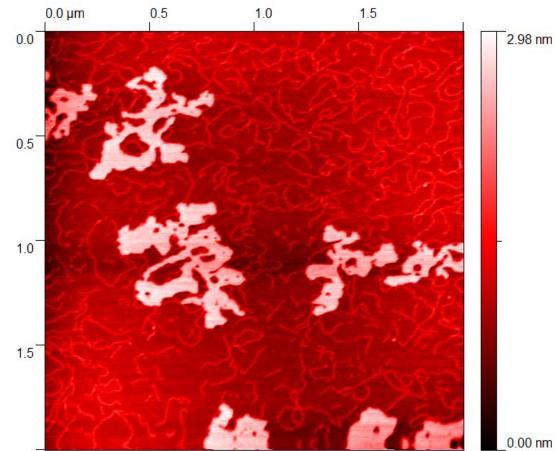
3-(Ethoxydimethylsilyl)propylamine APDMES



4.5 nm

#### Self-polymerization

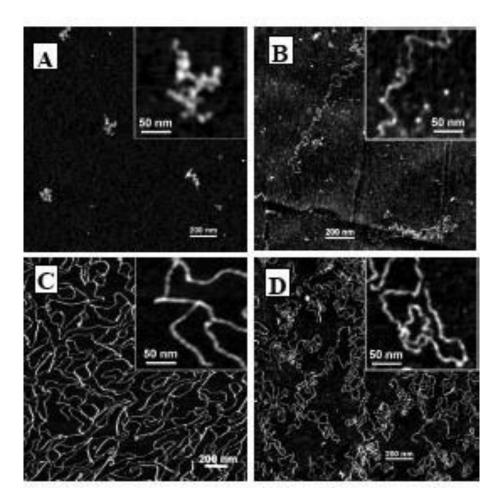
#### examples



#### **B. DNA on HOPG**

# Adsorption of long chain double-sided ions $(C_{16}/C_{18})$

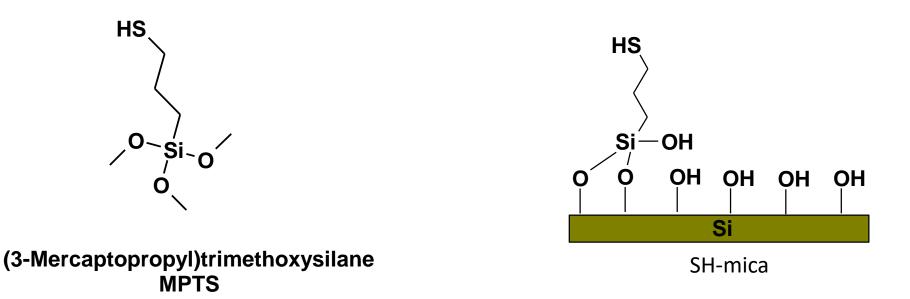
# HOPG H<sub>2</sub>N H<sub>2</sub>N H<sub>2</sub>N H<sub>2</sub>N $H_2N$ HOPG

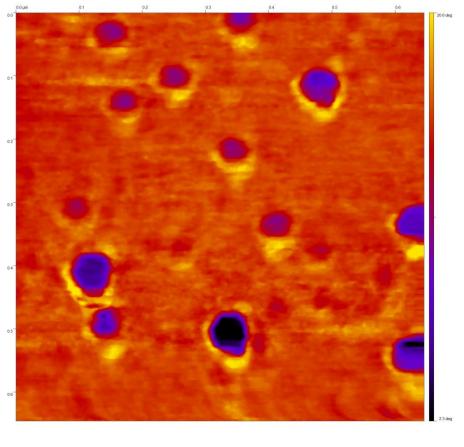


#### 3. Nanoparticles

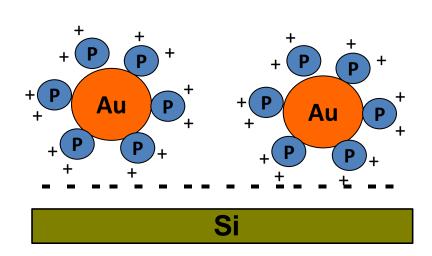
**Substrates** for immobilization: **mica** / **HOPG** (smooth surfaces), also gold, glass in selected cases.

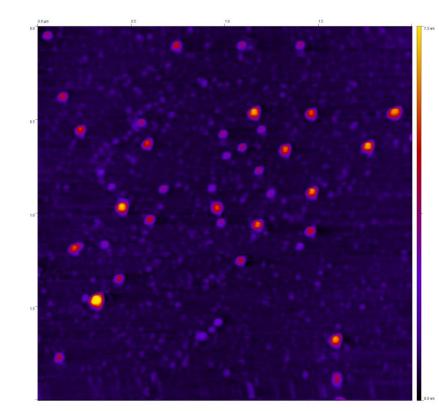
*Example*: gold nanoparticles (AuNP) mercapto-silanized mica (SH-mica):





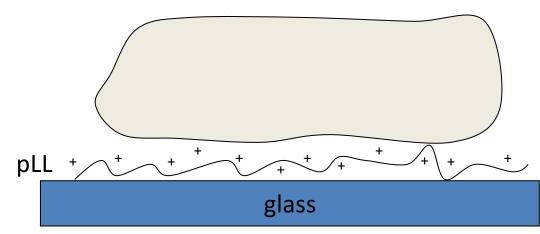
Gold nanoparticles (AuNP) conjugated with protein molecules: protein = immobilization bridge



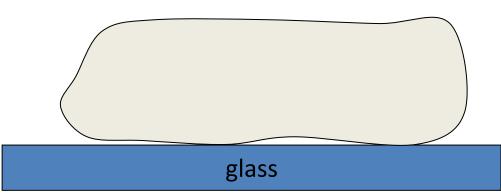


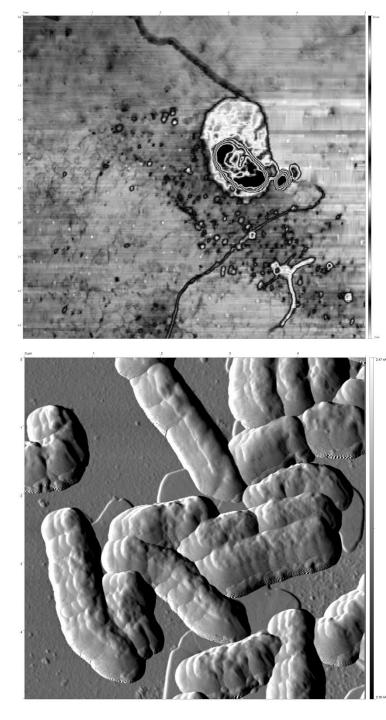
#### 3. Bacteria, spores

Protein adhesive layer, i.e. pLL (poly-L-lysine  $\rightarrow$  introducing positive charge)



#### Standard coating on glass

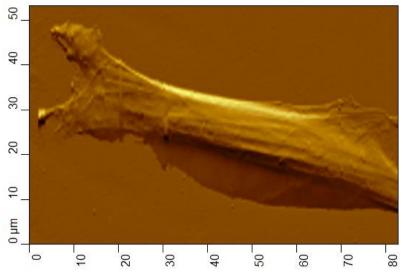


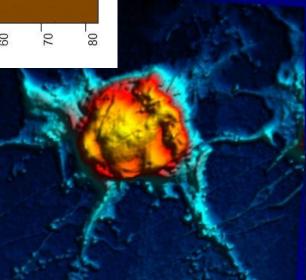


#### 5. Eukaryotic cells

#### A. Standard culturing on polystyrene dishes

Adhesive protein layers usually takes place (i.e. pLL, RGD adhesion factors, fibronectin, etc.)









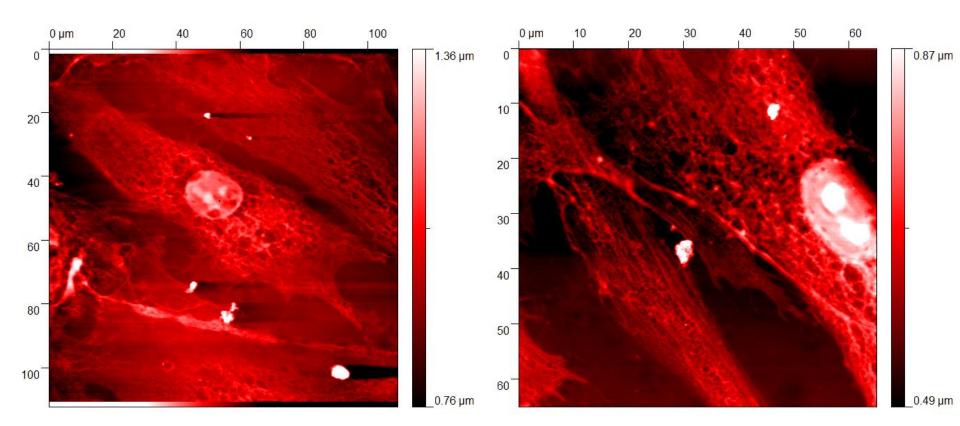
Cell culturing equipment



BioAFM incl. Petri dish heater for in-vitro imaging of cell cultures

#### **B. Fixation agents**

- Adhesion of cells out of incubator (37°C, 5% CO<sub>2</sub>) is mostly problematic
- Allows study of cells in long term periods after removal from incubator
- Cell wall destruction
- Example: EtOH, acetic acid, paraformaldehyde, glutardialdehyde











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#### Thank you for your attention!