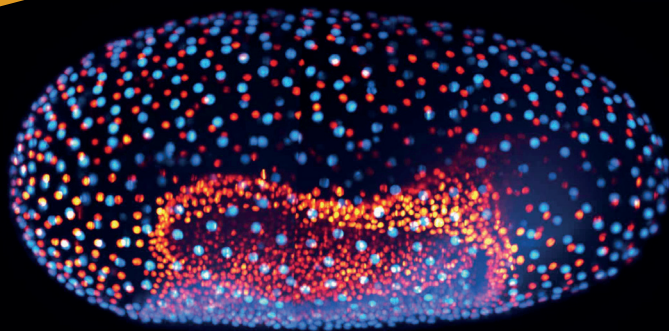


EMBO
Practical Course



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Light sheet microscopy

15 – 26 August 2022 | Brno, Czech Republic

Abstract Book

meetings.embo.org/event/22-light-sheet-micro

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EMBO
Practical Course

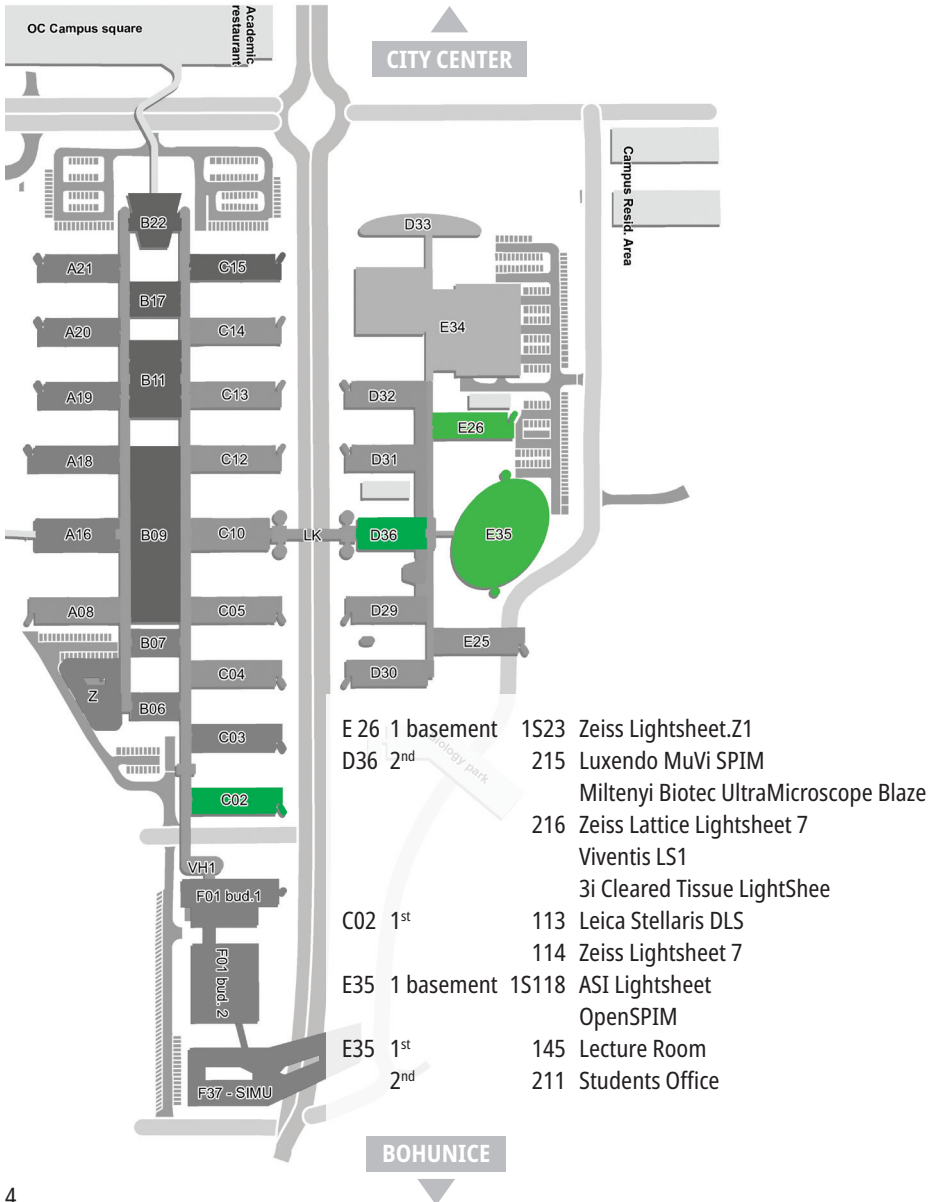
Light sheet microscopy

15 – 26 August 2022 | Brno, Czech Republic

Abstract Book



GENERAL INFORMATION



WELCOME

Dear course participant,

Welcome to the EMBO Practical Course on Light Sheet Microscopy at the Central European Institute of Technology CEITEC in Brno. This course is the continuation of a successful series of EMBO courses held at MPI-CBG in Dresden. With the core group of organizers, Emmanuel Reynaud and Jan Peychl, we tried to foster the adoption of the light sheet technology in life sciences over the past decade. Light sheet microscopy is now an established technology that enables imaging of large biological specimens with minimal photo-damage and at unprecedented speeds. A brief look at scientific literature suggests that light sheet technology opened up new avenues to study cell biological, neurobiological and developmental processes across multiple scales.

The challenges associated with light-sheet imaging remain the same; need for new paradigms in sample preparation, throughput of long term time lapse imaging and the enormity of the data that the microscopes generate and that need to be analyzed computationally. At CEITEC, we will try to show you how to tackle these challenges. In comparison with previous courses, we will put more emphasis on imaging of plant samples and photomanipulation of the imaged specimens. The course will, as before, revolve around your samples, your scientific questions and your ideas.

We are looking forward to an exciting, intense and hopefully rewarding two weeks of light sheet microscopy in the heart of Moravia. Brno is the place where Mendel did his groundbreaking genetic experiments and nowadays it is a home of thriving academic/ industrial ecosystem developing and producing state-of-the-art electron microscopes. Bringing light sheet imaging to this environment has a potential to combine the two technologies for the betterment of science and society.

Pavel Tomancak



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Useful Information



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Phone: 158



Phone: 150



Phone: 112

Organizational Information

Should you have any questions regarding the course itself, you might need help with food & accommodation, logistics, or perhaps you seek advice on what to see in Brno - please, do not hesitate to contact our organizational team. They are here to support you and make things smooth.

Please keep your badges with you throughout the course inside the Institute so that you can be identified when asked.



Wifi
ssid: MUNIguests
password: MUNIguests

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MON 22.8	TUE 23.8	WED 24.8	THU 25.8	FRI 26.8		



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	TUE 23.8					



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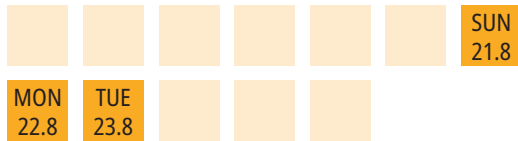
jozef.samaj@upol.cz

	TUE 23.8	WED 24.8	THU 25.8			



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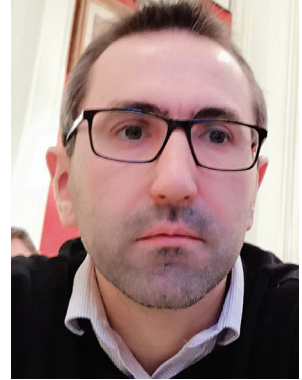
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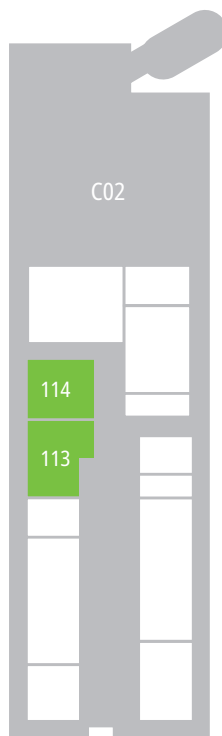
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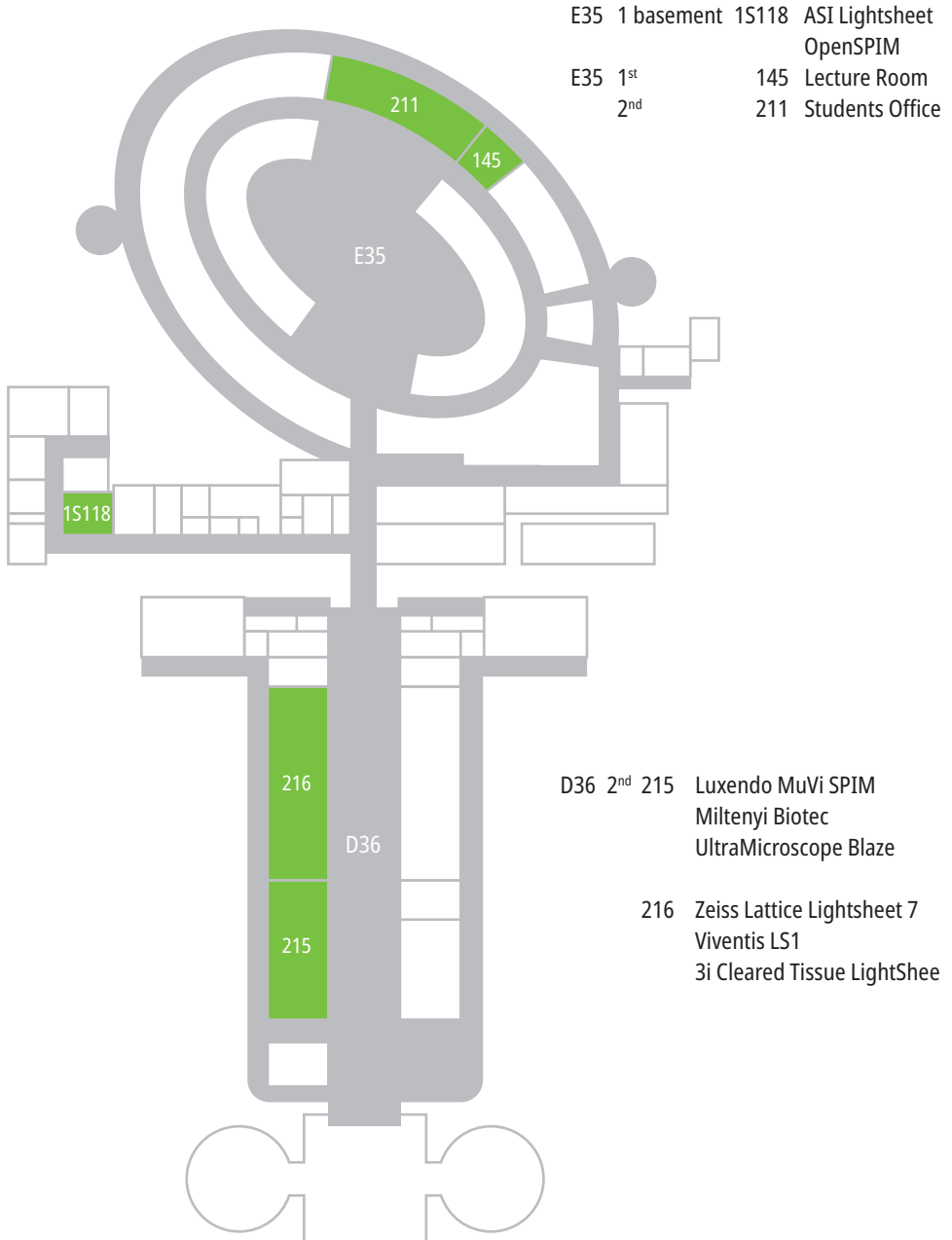
INSTRUMENT LOCATIONS

E 26 1 basement 1S23 Zeiss Lightsheet.Z1



C02 1st 113 Leica Stellaris DLS
114 Zeiss Lightsheet 7





INSTRUMENTS



Lattice Lightsheet 7



Illumination

Illumination: 13.3× / NA 0.4

Detection: 44.83× / NA 1.0

Beam shaping by cylinder lens and spatial light modulator (SLM)

Pre-defined Sinc3 beams with length

[μm]×thickness [nm]:

- 15×550 (w/ side lobes) & 15×650 (w/o side lobes)
- 30×700 (w/ side lobes) & 30×1,000 (w/o side lobes)
- 100×1,400 (w/ side lobes) & 100×1,800 (w/o side lobes)

Excitation

LED (white & red) for transmitted light Laser (488 nm, 561 nm, 640 nm) for reflected light and epi-fluorescence

Detection

Hamamatsu ORCA-Fusion

Filters

Emission Filter Camera 1

- BP 570-620 + LP 655
- BP 495-550 + LP 655
- LBF 405/488/561/642

- ND filter
- Empty
- BP 495-570
- LP 570

Emission Filter Camera 2

- BP 570-610 IR+
- Empty
- BP 495-550 + BP 570-620
- BP 500-550 IR+

Secondary Beam Splitter

- Plate
- LP 565
- LP 640
- Empty

Sample types

- Sample carrier frame Dish 35: for 35 mm cell culture dishes
- Sample carrier frame Dish 35...40: for 35 – 40 mm cell culture dishes
- Sample carrier frame Slide: for slides 26 mm × 76 mm; also suitable for multi-well glass-bottom slides 26 mm × 76 mm
- Sample carrier frame Chamber slide: for LabTekR chambers 25 mm × 57 mm; also suitable for multi-well glass-bottom slides 25 mm×57 mm
- Sample carrier frame Multiwell: for multiwell microplates 85.48×127.76 mm

Incubation

yes

Special features

Location

D36/216



Lightsheet 7



Illumination

Detection optics:

Water immersion:

- 5x / 0.16 foc (WD = 5.1 mm)
- 10x / 0.5 foc (WD = 3.7 mm)
- 20x / 1.0 foc (WD = 2.4 mm)

Clearing:

- Fluar 2.5x / 0.12 M27 (WD = 8.7 mm)
- 5x / 0.16 foc, n=1.33-1.58 (WD = 5.1 mm)
- Clr Plan-Neofluar 20x / 1.0 Corr nd=1.45 (WD = 5.6 mm)
- Clr Plan-Neofluar 20x / 1.0 Corr nd=1.53 (WD = 6.4 mm)

Illumination optics:

- 5x / 0.1 foc
- 10x / 0.2 foc

Excitation

405 nm, 488 nm, 561 nm, 638 nm

Detection

2x PCO edge 4.2 sCMOS camera, 1920 × 1920;
pixel size 6.5 μm × 6.5 μm

Filters

DAPI-GFP, GFP-Cy3, GFP-mCherry, GFP-Cy5,
Cy3-Cy5

Sample types

- Water chamber (n=1.33)
- 20x Clearing chamber (n=1.35-1.58)
- Large sample chamber (n=1.33-1.58)
- Translucence chamber

Incubation

yes

Special features

Location

C02/114



Lightsheet.Z1



Illumination

Dual-sided light sheet illumination with pivoting option, Illumination objectives: 5x/0.1 10/0.2

Excitation

405nm, 488nm, 561nm, 638nm

Detection

Single/dual-sided detection, 5x/0.16, 10x/0.5, 20x/1.0, 40x/1.0, camera 2x PCO Edge

Filters

LP 490, BP 420-470, BP 505-545, LP 510, BP 575-615, LP 560, LP 585, LP 660

Sample types

Sample chamber for live samples, agarose embedding holders. Sample chamber and optics designed for aqueous media (RI 1.33-). Short term (fast acquisition) and long term live imaging, 3D reconstruction

Incubation

yes

Special features

Location

E26/1S23



Stellaris DLS



Illumination

HC PL FLUOTAR 5x/0,15 IMM DLS
HC FLUOTAR L 25x/0,95 W DLS
HC FLUOTAR L 16x/0,60 IMM CORR DLS

Excitation

405, white pulsed laser 440 – 790nm

Detection

Hamamatsu ORCA

Filters

Sample types

Incubation

OKOLab incubation onstage

Special features

Location

C02/113



Miltenyi Biotec

UltraMicroscope Blaze



Illumination

Uni- and bi-directional, Objective lenses
1.1x, 4x, 12x

Excitation

Laser beam combiner: max. 5 laser lines
(405, 488, 561, 639, 785 nm), 50 mW–100 mW
per diode

Supercontinuum white light laser
460–800 nm, 1–3 mW/nm

Detection

4.2 Megapixel sCMOS camera
5.5 Megapixel sCMOS camera

Filters

Seven filters \varnothing 43 mm

Sample types

Cuvette for UltraMicroscope Blaze (standard
sample chamber)

XXL chamber for UltraMicroscope Blaze

Incubation

no

Special features

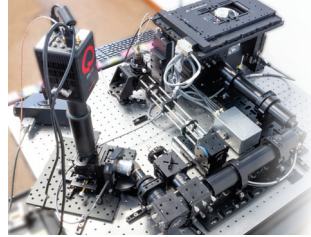
Location

D36/215



APPLIED SCIENTIFIC
INSTRUMENTATION

Lightsheet



Illumination

static sheet through detection objective

Excitation

488/561/640nm

Detection

40x/1.1 WI objective with correction collar
in inverted geometry, single sCMOS camera
with total magnification 67x

Filters

matching excitation

Sample types

aqueous live or fixed

Incubation

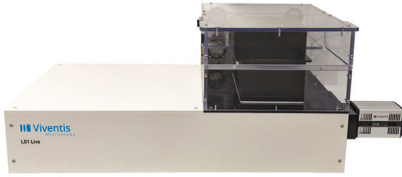
temperature and gas

Special features

fast volumetric imaging, compatible
with multi-well plates

Location

E35/1S118



Illumination

Scanned gaussian light sheet of variable thickness between 2.2 to 4.4 μm FWHM. Illumination with two 10X 0.2 NA

Excitation

488nm 60mW, 561 50mW, 638 100mW

Detection

25X 1.1 NA - field of view up to 900 μm .

Filters

Single band pass GFP // Double band pass GFP/mCherry(RFP) // Triple band pass GFP/mCherry(RFP)/FarRed

Sample types

Live sample or fix/expanded cleared sample in water-based solution (refractive index around 1.33).

Particularly suitable for organoids/spheroids/gastruloids and embryos imaging (i.e. mouse, c.elegans, zebrafish). Size limited by FOV of the microscope but tiling is possible during acquisition.

Incubation

Temperature controlled with heating above ambient temperature and CO_2 and O_2 controlled.

Special features

Sample is mounted in a dedicated FEP bottom dished where sample can be placed from top without need of embedding. Medium exchange is possible during acquisition.

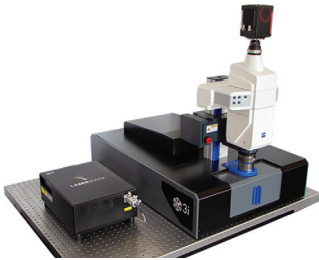
Location

D36/216



3i

Cleared Tissue
LightSheet



Illumination

dual-sided scanned infinity beam
two sets of excitation objectives
5x/0.14NA
10x/0.28NA

Excitation

488nm
561nm
640nm

Detection

1.0x/0.25NA
1.5x0.37NA
Hamamatsu Orca Flash 4.0 v3

Filters

Quad band-pass 446/523/600/677

Sample types

cleared, any type of clearing media

Incubation

heating

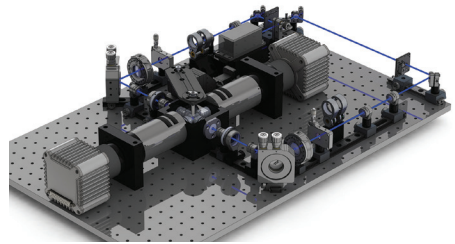
Special features

Location

D36/216



OpenSPiM OpenSPiM



Illumination

Dual-sided illumination and two detection axes

40x water-dipping objective, N.A 0.80

Excitation

Two laser lines (488 and 561) coming from a multiple wavelength laser system (Stradus VersaLase)

Detection

Two Andor sCMOS Neo 5.5 cameras

Filters

Sample types

Incubation

Yes

Special features

Location

E35/1S118



Illumination

Scanned Gaussian Beam light sheet microscope, Multi view SPIM - dual illumination / dual detection, Vertical sample mounting, Live and cleared sample imaging

Available objectives – exchange of objectives requires 15 minutes, therefore a pooling of applications is preferred minimizing the exchange of octagons/objectives

- 16X 0.8NA detection live samples
- 20X 1.0NA detection live samples
- 10X 0.5NA detection cleared samples
- 20X 1.0NA detection cleared samples

Excitation

405nm (>40mW), 488nm (>40mW), 561nm (>40mW), 642nm (>40mW)

Detection

3X sCMOS ORCA Flash4 3.0 Hamamatsu

Camera configuration:

- Left: Single camera detection
- Right: Dual camera detection – short & long wavelength
- 10X 1.0NA detection cleared samples

Filters

BP 418-462, LP 466, LP 498, BP497-554, LP 572, BP 580-627, LP 656, BP 655-704

Sample types

Live & fixed samples – up to 1.5 mm (l) × 1.5 mm (d) × 10 mm (h) – required imaging resolution dependent as objective combination may limit samples size

Cleared samples – up to 12 mm (l) × 15 mm (d) × 19 mm (h) – required imaging resolution dependent as objective combination may limit samples size

Incubation

Temperature control: 18 – 40 °C

CO₂ control: 0-20%

O₂ control: 0-20%

Humidity control

Special features

- Pivot scan for reduced shadowing effect
- Line mode acquisition
- Elongated light sheet for homogenous illumination
- Photo manipulation module for photo activation and ablation
- Dual color module

Location

D36/215

PROGRAMME

Lectures within the EMBO Course Light Sheet Microscopy

Building E35

Room 145

No registration necessary

Date/Time	Title	Speaker	Institution
15/08/2022 17:30–18:30	Introduction to Light Sheet Microscopy	Ernst Stelzer	Buchmann Institute for Molecular Life Sciences
16/08/2022 17:30–18:30	Light-sheet Imaging of Non-model Species	Tassos Pavlopoulos	IMBB-FORTH Greece
17/08/2022 9:00–10:00	Light Sheet Imaging of Mouse Embryos	Kate McDole	MRC LMB
18/08/2022 9:00–10:00	3D Image Analysis with Open Tools	Robert Haase	PoL Dresden
19/08/2022 9:00–10:00	Light Sheet Imaging of Insect Embryos	Akanksha Jain	ETH Zurich in Basel
22/08/2022 9:00–10:00	Light Sheet Imaging of Organoids	Gopi Shah	EMBL Barcelona
23/08/2022 9:00–10:00	Light Sheet Imaging of Bacterial Biofilms and Viruses	Pavel Plevka	CEITEC, Brno
24/08/2022 9:00–10:00	Light Sheet Imaging of Plants	Jozef Samaj	Palacky University Olomouc
25/08/2022 9:00–10:00	Light Sheet Imaging of Non-model Species	Tassos Pavlopoulos	IMBB-FORTH Greece



**SEE YOU
LATER**